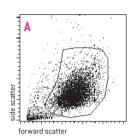


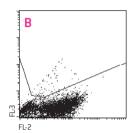
Amaxa® Cell Line Nucleofector® Kit V

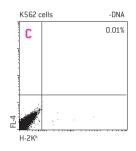
For K562 [DSMZ ACC10, cryopreserved]

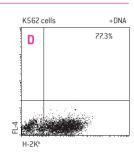
Human chronic myeloid leukemia cell line; round large single cells in suspension

Example for Nucleofection® of K562 cells









K562 cells (DSMZ ACC10) were transfected with the Cell Line Nucleofector® Kit V, Program T-003 and a plasmid encoding the mouse MHC class I heavy chain molecule H-2Kk. Cells were stained 5 hours post Nucleofection® with a PE-coupled antibody directed against H-2Kk and analyzed by flow cytometry. K562 cells were gated according to forward/side scatter (A). Dead cells were excluded by staining with propidium iodide and gating (B). H-2Kk expression is shown after Nucleofection® without (C) and with plasmid DNA (D).



Average transfection efficiency of K562 cells. K562 cells (DSMZ ACC10) were transfected with program T-003 and a plasmid encoding the enhanced green fluorescent protein eGFP. Cells were analyzed 5 and 24 hours post Nucleofection® by flow cytometry.

Product Description

Cat. No.		VCA-1003
Size (reactions)		25
Cell Line Nucleofector® Solution V		2.25 ml (2.05 ml + 10% overfill)
Supplement		0.5 ml (0.45 ml + 10% overfill)
pmaxGFP® Vector (0.5 µg/µl in 10 mM Tris pH 8.0)		30 µg
Certified cuvettes		25
Plastic pipettes		25
Storage and stability	Store Nucleofector® Solu	ution, Supplement and pmaxGFP® Vector at 4°C. For long-term storage,

Store Nucleofector® Solution, Supplement and pmaxGFP® Vector at 4°C. For long-term storage, pmaxGFP® Vector is ideally stored at -20°C. The expiration date is printed on the solution box. Once the Nucleofector® Supplement is added to the Nucleofector® Solution it is stable for three months at 4°C.

Optimized Protocol for K562 Cells [DSMZ]

Required Material

Note

Please make sure that the entire supplement is added to the Nucleofector® Solution. The ratio of Nucleofector® Solution to supplement is 4.5:1. For a single reaction use 82 μ l of Nucleofector® Solution plus 18 μ l of supplement to make 100 μ l of total reaction volume.

- Nucleofector® Device
- Supplemented Nucleofector® Solution at room temperature
- Supplied certified cuvettes
- Supplied plastic pipettes
- Supplied pmaxGFP® Vector
- Substrate of interest, highly purified, preferably by using endotoxin-free kits; A260: A280 ratio should be at least 1.8
- 12-well culture dish or culture system of your choice
- Culture medium: RPMI 1640 [Lonza; Cat. No. BE12-167F] supplemented with 2 mM UltraGlutamine I [Lonza, Cat. No. BE17-605E/U1], 100 μg/ml streptomycin, 100 U/ml penicillin and 10% fetal calf serum [FCS]
- Prewarm appropriate volume of culture medium to 37°C (2 ml per sample)
- Appropriate number of cells (1 x 10^6 2 x 10^6 cells per sample; lower or higher cell numbers may influence transfection results)

1. Pre Nucleofection®

Cell culture recommendations

- 1.1 Passage cells after reaching $1 5 \times 10^5$ cells/ml
- 1.2 Seed out $3 5 \times 10^5$ cells/ml
- 1.3 Subculture 2 days before Nucleofection®
- 1.4 Optimal cell density for Nucleofection $^{\circ}$: 2 5 x 10 $^{\circ}$ cells/ml. Higher cell densities may cause lower Nucleofection $^{\circ}$ Efficiencies

Optimized Protocol for K562 Cells [DSMZ]

2. Nucleofection®

One Nucleofection® Sample contains

 $1 - 2 \times 10^{6}$ cells

 $1-5 \mu g$ plasmid DNA (in $1-5 \mu l$ H₂0 or TE) or $2 \mu g$ pmaxGFP® Vector or 30-300nM siRNA (3-30 pmol/sample)

100 µl Cell Line Nucleofector® Solution V

- 2.1 Please make sure that the entire supplement is added to the Nucleofector® Solution
- 2.2 Prepare 12-well plates by filling appropriate number of wells with 1.5 ml of supplemented culture media and pre-incubate/equilibrate plates in a humidified 37°C/5% CO₂ incubator
- 2.3 Count an aliquot of the cells and determine cell density
- 2.4 Centrifuge the required number of cells $(1 2 \times 10^6)$ cells per sample at 200xg for 10 minutes at room temperature. Remove supernatant completely
- 2.5 Resuspend the cell pellet carefully in 100 µl room-temperature Nucleofector® Solution per sample

Note Avoid leaving the cells in Nucleofector® Solution for extended periods of time (longer than 15 minutes), as this may reduce cell viability and gene transfer efficiency.

- 2.6 Combine 100 μ l of cell suspension with 2 μ g DNA, 2 μ g pmaxGFP® Vector or 30 nM 300 nM siRNA (3 30 pmol/sample) or other substrates
- 2.7 Transfer cell/DNA suspension into certified cuvette (sample must cover the bottom of the cuvette without air bubbles). Close the cuvette with the cap
- 2.8 Select the appropriate Nucleofector® Program T-003 (T-03 for Nucleofector® | Device)
- 2.9 Insert the cuvette with cell/DNA suspension into the Nucleofector® Cuvette Holder and apply the selected program by pressing the X-button
- 2.10 Take the cuvette out of the holder once the program is finished
- 2.11 Immediately add \sim 500 μ l of the pre-equilibrated culture medium to the cuvette and gently transfer the sample into the prepared 12-well plate (final volume 2 ml media per well). Use the supplied pipettes and avoid repeated aspiration of the sample

3. Post Nucleofection®

3.1 Incubate the cells in humidified 37°C/5% $\rm CO_2$ incubator until analysis. Gene expression or down regulation, respectively, is often detectable after only 4 - 8 hours

Additional Information

For an up-to-date list of all Nucleofector® References, please refer to: www.lonza.com/nucleofection-citations

For more technical assistance, contact our Scientific Support Team:

USA/Canada Europe and Rest of World

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Lonza Cologne AG 50829 Cologne, Germany

Please note that the Amaxa® Nucleofector® Technology is not intended to be used for diagnostic purposes or for testing or treatment in humans.

The Nucleofector® Technology, comprising Nucleofection® Process, Nucleofector® Device, Nucleofector® Solutions, Nucleofector® 96-well Shuttle® System and 96-well Nucleocuvette® plates and modules is covered by patent and/or patent-pending rights owned by Lonza Cologne AG.

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