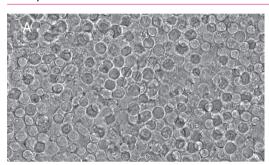
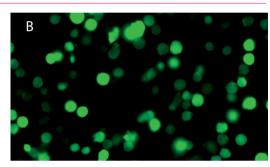
# Amaxa® Cell Line Nucleofector® Kit V

## For 32 D

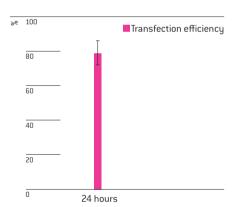
Mouse bone marrow; lymphoblastoid cells

#### Example for Nucleofection® of A-431 cells of 32 D cells





32 D cells were transfected with the Cell Line Nucleofector® Kit V, Program E-032 and 2 µg of pmaxGFP® Vector. Cells were analyzed 24 hours post Nucleofection® using light (A) and fluorescence microscopy (B).



Average transfection efficiency of 32 D cells. 32 D cells were transfected with program E-032 and 2 µg of pmaxGFP® Vector. Cells were analyzed 24 hours post Nucleofection® by flow cytometry. Cell viability is around 61% 24 hours post Nucleofection®.

## **Product Description**

Cat. No.	VCA-1003
Size (reactions)	25
Cell Line Nucleofector® Solution V	2.25 ml (2.05 ml + 10% overfill)
Supplement	0.5 ml (0.45 ml + 10% overfill)
pmaxGFP® Vector (0.5 μg/μl in 10 mM Tris pH 8.0)	30 µg
Certified cuvettes	25
Plastic pipettes	25

Storage and stability

Store Nucleofector® Solution, Supplement and pmaxGFP® Vector at 4°C. For long-term storage, pmaxGFP® Vector is ideally stored at -20°C. The expiration date is printed on the solution box. Once the Nucleofector® Supplement is added to the Nucleofector® Solution it is stable for three months at 4°C.

## Optimized Protocol for 32 D

## **Required Material**

Note

Please make sure that the entire supplement is added to the Nucleofector® Solution. The ratio of Nucleofector® Solution to supplement is 4.5:1. For a single reaction use  $82\,\mu$ l of Nucleofector® Solution plus  $18\,\mu$ l of supplement to make  $100\,\mu$ l of total reaction volume.

- Nucleofector® Device; Software requirements: version V2.3 or higher for Nucleofector® I Device; version
   S3-4 or higher for Nucleofector® II Device
- Supplemented Nucleofector® Solution at room temperature
- Supplied certified cuvettes
- Supplied plastic pipettes
- Supplied pmaxGFP® Vector
- Substrate of interest, highly purified, preferably by using endotoxin-free kits; A260: A280 ratio should be at least 1.8
- 12-well culture dish or culture system of your choice
- Culture medium: complete growth medium: RPMI 1640 medium with 2 mM L-glutamine adjusted to contain 1.5 g/L sodium bicarbonate, 4.5 g/L glucose, 10 mM HEPES, and 1.0 mM sodium pyruvate supplemented with 10% heat-inactivated fetal bovine serum, 10% mouse Interleukin-3 culture supplement [Becton Dickinson Cat. No. 354040]
- Prewarm appropriate volume of culture medium to 37°C (1.5 ml per sample)
- Appropriate number of cells (1 x  $10^6$  cells per sample; lower or higher cell numbers may influence transfection results)

#### 1. Pre Nucleofection®

#### Cell culture recommendations

- 1.1 Replace media every 2 3 days
- 1.2 Passage cells every 2 3 days.
- 1.3 Seed out  $2 \times 10^5 1 \times 10^6$  viable cells/ml
- 1.4 Subculture 1 2 days before Nucleofection®
- 1.5 Optimal confluency for Nucleofection®: 75%. Higher cell densities may cause lower Nucleofection® Efficiencies

#### For detaching cells:

1.6 Scrape off the attached cells and transfer along with floating cells into new flasks

## Optimized Protocol for 32 D

#### 2. Nucleofection®

#### One Nucleofection® Sample contains

1 x 106 cells

2  $\mu$ g plasmid DNA (in 1 – 5  $\mu$ l H $_2$ 0 or TE) or 2  $\mu$ g pmaxGFP® Vector or 30 – 300nM siRNA (3 – 30 pmol/sample)

100 µl Cell Line Nucleofector® Solution V

- 2.1 Please make sure that the entire supplement is added to the Nucleofector® Solution
- 2.2 Prepare 12-well plates by filling appropriate number of wells with 1 ml of supplemented culture media and pre-incubate/equilibrate plates in a humidified 37°C/5% CO<sub>2</sub> incubator
- 2.3 Count an aliquot of the cells and determine cell density
- 2.4 Centrifuge the required number of cells (1 x  $10^6$  cells per sample) at 90xg for 10 minutes at room temperature. Remove supernatant completely
- 2.5 Resuspend the cell pellet carefully in 100 µl room-temperature Nucleofector® Solution per sample

Note Avoid leaving the cells in Nucleofector® Solution for extended periods of time (longer than 15 minutes), as this may reduce cell viability and gene transfer efficiency

- 2.6 Combine 100  $\mu$ l of cell suspension with 2  $\mu$ g DNA, 2  $\mu$ g pmaxGFP® Vector or 30 nM 300 nM siRNA (3 30 pmol/sample) or other substrates
- 2.7 Transfer cell/DNA suspension into certified cuvette (sample must cover the bottom of the cuvette without air bubbles). Close the cuvette with the cap
- 2.8 Select the appropriate Nucleofector® Program E-032 (E-32 for Nucleofector® | Device)
- 2.9 Insert the cuvette with cell/DNA suspension into the Nucleofector® Cuvette Holder and apply the selected program by pressing the X-button
- 2.10 Take the cuvette out of the holder once the program is finished
- 2.11 Immediately add  $\sim$  500  $\mu$ l of the pre-equilibrated culture medium to the cuvette and gently transfer the sample into the prepared 12-well plate (final volume 1.5 ml media per well). Use the supplied pipettes and avoid repeated aspiration of the sample

## 3. Post Nucleofection®

3.1 Incubate the cells in humidified 37°C/5%  $\rm CO_2$  incubator until analysis. Gene expression or down regulation, respectively, is often detectable after only 4 - 8 hours

Note For detaching the adherent cells post Nucleofection® in 12-well plates, we recommend incubating the adherent cells in ice-cold PBS for 5 – 10 min and then rinsing the well.

## **Additional Information**

For an up-to-date list of all Nucleofector® References, please refer to: www.lonza.com/nucleofection-citations

#### For more technical assistance, contact our Scientific Support Team:

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