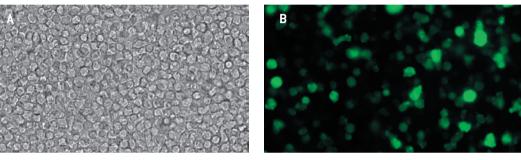
Lonza

Amaxa[®] Cell Line Nucleofector[®] Kit T

For NALM-6 [DSMZ ACC128, cryopreserved]

Human B cell precursor leukemia; single, small, round cells in suspension

Example for Nucleofection® of NALM-6 cells



NALM-6 cells [DSMZ ACC128] were transfected with the Cell Line Nucleofector® Kit T, Program C-005 and 2 µg of pmaxGFP® Vector. Cells were analyzed 24 hours post Nucleofection® using light (A) and fluorescence microscopy (B).



Average transfection efficiency of NALM-6 cells. NALM-6 cells [DSMZ ACC128] were transfected with program C-005 and 2 μ g of pmaxGFP® Vector. Cells were analyzed 24 hours post Nucleofection® by flow cytometry. Cell Viability (% PI negative) is around 87% 24 hours post Nucleofection®.

Product Description

Cat. No.		VCA-1002
Size (reactions)		25
Cell Line Nucleofector [®] Solution T		2.25 ml (2.05 ml + 10% overfill)
Supplement		0.5 ml (0.45 ml + 10% overfill)
pmaxGFP® Vector (0.5 µg/µl in 10 mM Tris pH 8.0)		30 µg
Certified cuvettes		25
Plastic pipettes		25
Storage and stability	Store Nucleofector® Sol	lution, Supplement and pmaxGFP [®] Vector at 4°C. For long-term storage,
	pmaxGFP® Vector is ideal	Ily stored at -20°C. The expiration date is printed on the solution box. Once the
	Nucleofector [®] Suppleme	nt is added to the Nucleofector® Solution it is stable for three months at 4°C.

Required Material

Note

Please make sure that the entire supplement is added to the Nucleofector[®] Solution. The ratio of Nucleofector[®] Solution to supplement is 4.5:1. For a single reaction use 82 µl of Nucleofector[®] Solution plus 18 µl of supplement to make 100 µl of total reaction volume.

- Nucleofector[®] Device
- Supplemented Nucleofector® Solution at room temperature
- Supplied certified cuvettes
- Supplied plastic pipettes
- Supplied pmaxGFP[®] Vector
- Substrate of interest, highly purified, preferably by using endotoxin-free kits; A260: A280 ratio should be at least 1.8
- 12-well culture dish or culture system of your choice
- Culture medium: RPMI 1640, 90% [Lonza; Cat. No. BE12-167F], fetal bovine serum, 10%
- Prewarm appropriate volume of culture medium to 37°C (1.5 ml per sample)
- Appropriate number of cells (2 x 10⁶ cells per sample; lower or higher cell numbers may influence transfection results)

1. Pre Nucleofection®

Cell culture recommendations

- 1.1 Replace media every 3 4 days
- 1.2 Passage cells 2 times a week. A subcultivation ratio of 1:2 to 1:4 is recommended
- 1.3 Maintain cultures between $1-2\,x\,10^{6}\,cells/ml$
- 1.4 Seed out 1 x 10 6 cells/ml. Minimum density of 5 x 10 5 cells/ml
- 1.5 Subculture 3 days before Nucleofection®

2. Nucleofection®

One Nucleofection® Sample contains

2 x 10⁶ cells 2 μg plasmid DNA (in 1 – 5 μl H₂O or TE) or 2 μg pmaxGFP® Vector or 30 – 300nM siRNA (3 – 30 pmol/sample) 100 μl Cell Line Nucleofector® Solution T

- 2.1 Please make sure that the entire supplement is added to the Nucleofector[®] Solution
- 2.2 Prepare 12-well plates by filling appropriate number of wells with 1 ml of supplemented culture media and pre-incubate/equilibrate plates in a humidified 37°C/5% CO₂ incubator
- 2.3 Count an aliquot of the cells and determine cell density
- 2.4 Centrifuge the required number of cells (2 x 10⁶ cells per sample) at 90xg for 10 minutes at room temperature. Remove supernatant completely
- 2.5 Resuspend the cell pellet carefully in 100 µl room-temperature Nucleofector® Solution per sample

Note Avoid leaving the cells in Nucleofector[®] Solution for extended periods of time (longer than 15 minutes), as this may reduce cell viability and gene transfer efficiency.

- 2.6 Combine 100 μ l of cell suspension with **2 \mug DNA**, 2 μ g pmaxGFP[®] Vector or **30 nM 300 nM** siRNA (3 30 pmol/sample) or other substrates
- 2.7 Transfer cell/DNA suspension into certified cuvette (sample must cover the bottom of the cuvette without air bubbles). Close the cuvette with the cap
- 2.8 Select the appropriate Nucleofector® Program C-005 (C-05 for Nucleofector® | Device)
- 2.9 Insert the cuvette with cell/DNA suspension into the Nucleofector[®] Cuvette Holder and apply the selected program by pressing the X-button
- 2.10 Take the cuvette out of the holder once the program is finished
- 2.11 Immediately add ~500 µl of the pre-equilibrated culture medium to the cuvette and gently transfer the sample into the prepared 12-well plate (final volume 1.5 ml media per well). Use the supplied pipettes and avoid repeated aspiration of the sample

3. Post Nucleofection®

3.1 Incubate the cells in humidified 37°C/5% CO_2 incubator until analysis. Gene expression or down regulation, respectively, is often detectable after only 4 – 8 hours

Additional Information

For an up-to-date list of all Nucleofector® References, please refer to: www.lonza.com/nucleofection-citations

For more technical assistance, contact our Scientific Support Team:

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Please note that the Amaxa® Nucleofector® Technology is not intended to be used for diagnostic purposes or for testing or treatment in humans.

The Nucleofector[®] Technology, comprising Nucleofection[®] Process, Nucleofector[®] Device, Nucleofector[®] Solutions, Nucleofector[®] 96-well Shuttle[®] System and 96-well Nucleocuvette[®] plates and modules is covered by patent and/or patent-pending rights owned by Lonza Cologne AG.

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