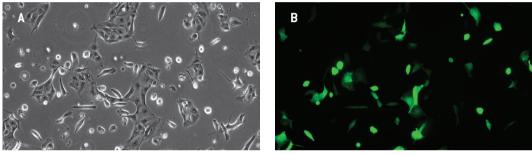
# Lonza

## Amaxa<sup>®</sup> Cell Line Nucleofector<sup>®</sup> Kit L

## For MDCK

Dog kidney; epithelial cells

#### Example for Nucleofection® of MDCK cells



MDCK cells were transfected with the Nucleofector<sup>®</sup> Kit L, Program A-024 and 5 µg of pmaxGFP<sup>®</sup> Vector. Cells were analyzed 24 hours post Nucleofection<sup>®</sup> using light (A) and fluorescence microscopy (B).



Average transfection efficiency of MDCK cells. MDCK cells were transfected with program A-024 and 5  $\mu$ g of pmaxGFP® Vector. Cells were analyzed 24 hours post Nucleofection® by flow cytometry. Cell Viability (%PI negative) is around 83% after 24 hours post Nucleofection®.

## **Product Description**

Cat. No.		VCA-1005
Size (reactions)		25
Cell Line Nucleofector® Solution L		2.25 ml (2.05 ml + 10% overfill)
Supplement		0.5 ml (0.45 ml + 10% overfill)
pmaxGFP® Vector (0.5 µg/µl in 10 mM Tris pH 8.0)		30 µg
Certified cuvettes		25
Plastic pipettes		25
Storage and stability	Store Nucleofector® Solu	ution, Supplement and pmaxGFP <sup>®</sup> Vector at 4°C. For long-term storage,
	pmaxGFP® Vector is ideall	ly stored at -20°C. The expiration date is printed on the solution box. Once the
	Nucleofector <sup>®</sup> Supplemer	nt is added to the Nucleofector® Solution it is stable for three months at 4°C.

## **Required Material**

#### Note

Please make sure that the entire supplement is added to the Nucleofector® Solution. The ratio of Nucleofector® Solution to supplement is 4.5 : 1. For a single reaction use 82 µl of Nucleofector® Solution plus 18 µl of supplement to make 100 µl of total reaction volume.

- Nucleofector<sup>®</sup> Device
- Supplemented Nucleofector® Solution at room temperature
- Supplied certified cuvettes
- Supplied plastic pipettes
- Supplied pmaxGFP<sup>®</sup> Vector
- Substrate of interest, highly purified, preferably by using endotoxin-free kits; A260 : A280 ratio should be at least 1.8
- 6-well culture dish or culture system of your choice
- For detaching cells: 2.5 mg/ml Trypsin and 1.0 mg/ml EDTA in PBS [Gibco; Cat. No. 15400-054: dilution 1:2] (10 15 min at 37°C) and supplemented culture media or PBS/0.5% BSA
- Culture medium: Minimum essential medium (Eagle) with 2 mM L-glutamine and Earle's BSS adjusted to contain 1.5 g/L sodium bicarbonate, 0.1 mM non-essential amino acids and 1.0 mM sodium pyruvate, 90%; fetal bovine serum, 10%
- Prewarm appropriate volume of culture medium to 37°C (1.5 ml per sample)
- Appropriate number of cells (5 x 10<sup>5</sup> cells per sample; lower or higher cell numbers may influence transfection results)

## 1. Pre Nucleofection®

#### Cell culture recommendations

- 1.1 Replace media every 2 3 days
- 1.2 Passage cells every 2 3 days into a new culture vessel. A subcultivation ratio of 1:3 to 1:5 is recommended
- 1.3 Do not passage more than 20 times
- 1.4 Use low spin centrifugation
- 1.5 Resuspend cells very gently
- 1.6 Subculture 2 3 days before Nucleofection®

#### Trypsinization

- 1.7 Remove media from the cultured cells and wash cells once with PBS; use at least same volume of PBS as culture media
- 1.8 For harvesting, incubate the cells ~10 − 15 minutes at 37°C with indicated trypsinization reagent (please see required material)
- 1.9 Neutralize trypsinization reaction with supplemented culture medium or PBS/0.5% BSA once the majority of the cells (>90%) have been detached

## 2. Nucleofection®

#### One Nucleofection® Sample contains

5 x 10⁵ cells

5 μg plasmid DNA (in 1 – 5 μl H<sub>2</sub>O or TE) or 2 μg pmaxGFP® Vector or 30 – 300nM siRNA <u>(3 – 30 pmol/sample)</u> 100 μl Cell Line Nucleofector® Solution L

- 2.1 Please make sure that the entire supplement is added to the Nucleofector® Solution
- 2.2 Prepare 6-well plates by filling appropriate number of wells with 1 ml of supplemented culture media and pre-incubate/equilibrate plates in a humidified 37°C/5% CO<sub>2</sub> incubator
- 2.3 Harvest the cells by trypsinization (please see 1.7 1.9)
- 2.4 Count an aliquot of the cells and determine cell density
- 2.5 Centrifuge the required number of cells (5 x 10<sup>5</sup> cells per sample) at 90xg for 10 minutes at room temperature. Remove supernatant completely
- 2.6 Resuspend the cell pellet carefully in 100 µl room-temperature Nucleofector® Solution per sample

## Note Avoid leaving the cells in Nucleofector<sup>®</sup> Solution for extended periods of time (longer than 15 minutes), as this may reduce cell viability and gene transfer efficiency.

- 2.7 Combine 100  $\mu$ l of cell suspension with **5**  $\mu$ g DNA, 2  $\mu$ g pmaxGFP<sup>®</sup> Vector or **30** nM **300** nM siRNA (3 30 pmol/sample) or other substrates
- 2.8 Transfer cell/DNA suspension into certified cuvette (sample must cover the bottom of the cuvette without air bubbles). Close the cuvette with the cap
- 2.9 Select the appropriate Nucleofector® Program: A-024 (A-24 for Nucleofector® | Device)
- 2.10 Insert the cuvette with cell/DNA suspension into the Nucleofector<sup>®</sup> Cuvette Holder and apply the selected program by pressing the X-button
- 2.11 Take the cuvette out of the holder once the program is finished
- 2.12 Immediately add ~500 µl of the pre-equilibrated culture medium to the cuvette and gently transfer the sample into the prepared 6-well plate (final volume 1.5 ml media per well). Use the supplied pipettes and avoid repeated aspiration of the sample

### 3. Post Nucleofection®

3.1 Incubate the cells in humidified 37°C/5%  $CO_2$  incubator until analysis. Gene expression or down regulation, respectively, is often detectable after only 4 – 8 hours

## Additional Information

For an up-to-date list of all Nucleofector® References, please refer to: www.lonza.com/nucleofection-citations

#### For more technical assistance, contact our Scientific Support Team:

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