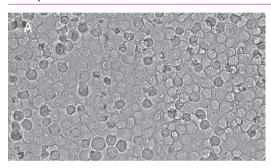
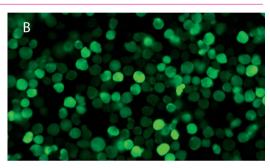
Amaxa® Cell Line Nucleofector® Kit V

For BA/F3 [DSMZ ACC-300 cryopreserved]

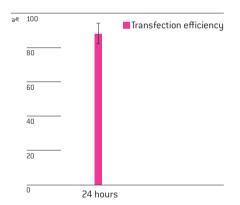
Mouse pro B cell; mostly single, round (some polymorph) cells in suspension (or occasionally in clumps)

Example for Nucleofection® of BA/F3 cells





BA/F3 cells (DSMZ ACC-300) were transfected with the Cell Line Nucleofector® Kit V, Program X-001 and 2 μg of pmaxGFP® Vector. Cells were analyzed 24 hours post Nucleofection® using light (A) and fluorescence microscopy (B).



Average transfection efficiency of BA/F3 cells. BA/F3 cells [DSMZ ACC-300] were transfected with program X-001 and 2 µg of pmaxGFP® Vector. Cells were analyzed 24 hours post Nucleofection® by flow cytometry. Cell viability [compared to non-transfected control] is around 80% 24 hours post Nucleofection®.

Product Description

Cat. No.	VCA-1003
Size (reactions)	25
Cell Line Nucleofector® Solution V	2.25 ml (2.05 ml + 10% overfill)
Supplement	0.5 ml (0.45 ml + 10% overfill)
pmaxGFP® Vector (0.5 μg/μl in 10 mM Tris pH 8.0)	30 µg
Certified cuvettes	25
Plastic pipettes	25

Storage and stability

Store Nucleofector® Solution, Supplement and pmaxGFP® Vector at 4°C. For long-term storage, pmaxGFP® Vector is ideally stored at -20°C. The expiration date is printed on the solution box. Once the Nucleofector® Supplement is added to the Nucleofector® Solution it is stable for three months at 4°C.

Optimized Protocol for BA/F3 Cell Line [DSMZ]

Required Material

Note Please make sure that the entire supplement is added to the Nucleofector® Solution. The ratio of Nucleofector® Solution to supplement is 4.5:1. For a single reaction use 82 µl of Nucleofector® Solution plus 18 µl of supplement to make 100 µl of total reaction volume.

- Nucleofector® Device; Software requirements: version V2.3 or higher for Nucleofector® I Device; version
 S3-4 or higher for Nucleofector® II Device
- Supplemented Nucleofector® Solution at room temperature
- Supplied certified cuvettes
- Supplied plastic pipettes
- Supplied pmaxGFP® Vector
- Substrate of interest, highly purified, preferably by using endotoxin-free kits; A260: A280 ratio should be at least 1.8
- 12-well culture dish or culture system of your choice
- Culture medium: RPMI 1640 [Lonza; Cat. No. BE12-167F], 90%; fetal bovine serum, 10%; UltraGlutamine I [Lonza; Cat. No. BE17-605E/U1], 1%; mouse IL-3 [BD; Cat. No. 354040], 10 ng/ml
- Prewarm appropriate volume of culture medium to 37°C (1.5 ml per sample)
- Appropriate number of cells (2 x 10^6 cells per sample; lower or higher cell numbers may influence transfection results)

1. Pre Nucleofection®

Cell culture recommendations

- 1.1 Replace media every 2 3 days
- 1.2 Passage cells every 2 3 days. A subcultivation ratio of 1:15 to 1:20 is recommended. Do not passage more than 25 times

Note At high density (> 2 x 10⁶ cells/ml), a cytokine-independent subclone may grow out relatively quickly.

- 1.3 Seed out $1 3 \times 10^5$ cells/ml in a T162 flask
- 1.4 Subculture 2 3 days before Nucleofection®

Optimized Protocol for BA/F3 Cell Line [DSMZ]

2. Nucleofection®

One Nucleofection® Sample contains

 2×10^6 cells

 $2 \mu g \text{ plasmid DNA (in } 1-5 \mu l \text{ H}_2\text{O or TE) or } 2 \mu g \text{ pmaxGFP}^{\circ} \text{ Vector or } 30-300 \text{nM siRNA } (3-30 \text{ pmol/sample})$

100 µl Cell Line Nucleofector® Solution V

- 2.1 Please make sure that the entire supplement is added to the Nucleofector® Solution
- 2.2 Prepare 12-well plates by filling appropriate number of wells with 1 ml of supplemented culture media and pre-incubate/equilibrate plates in a humidified 37°C/5% CO₂ incubator
- 2.3 Count an aliquot of the cells and determine cell density
- 2.4 Centrifuge the required number of cells (2×10^6 cells per sample) at 90xg for 10 minutes at room temperature. Remove supernatant completely
- 2.5 Resuspend the cell pellet carefully in 100 µl room-temperature Nucleofector® Solution per sample

Note Avoid leaving the cells in Nucleofector® Solution for extended periods of time (longer than 15 minutes), as this may reduce cell viability and gene transfer efficiency.

- 2.6 Combine 100 μ l of cell suspension with 2 μ g DNA, 2 μ g pmaxGFP® Vector or 30 nM 300 nM siRNA (3 30 pmol/sample) or other substrates
- 2.7 Transfer cell/DNA suspension into certified cuvette (sample must cover the bottom of the cuvette without air bubbles). Close the cuvette with the cap
- 2.8 Select the appropriate Nucleofector® Program X-001 [X-01 for Nucleofector® | Device]
- 2.9 Insert the cuvette with cell/DNA suspension into the Nucleofector® Cuvette Holder and apply the selected program by pressing the X-button
- 2.10 Take the cuvette out of the holder once the program is finished
- 2.11 Immediately add \sim 500 μ l of the pre-equilibrated culture medium to the cuvette and gently transfer the sample into the prepared 12-well plate (final volume 1.5 ml media per well). Use the supplied pipettes and avoid repeated aspiration of the sample

3. Post Nucleofection®

3.1 Incubate the cells in humidified 37°C/5% $\rm CO_2$ incubator until analysis. Gene expression or down regulation, respectively, is often detectable after only 4 – 8 hours

Optimized Protocol for BA/F3 Cell Line [DSMZ]

Additional Information

For an up-to-date list of all Nucleofector® References, please refer to: www.lonza.com/nucleofection-citations

For more technical assistance, contact our Scientific Support Team:

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